

Supplementary Information

Modular Nucleic Acid Assembled p/MHC Microarrays for Multiplexed Sorting of Antigen-Specific T Cells

Gabriel A. Kwong^{1,2}, Caius G. Radu^{1,6}, Kiwook Hwang^{1,4}, Chengyi J. Shu^{1,6}, Chao Ma^{1,3}, Richard C. Koya^{1,8}, Begonya Comin-Anduix^{1,8}, Sine Reker Hadrup⁵, Ryan C. Bailey^{1,4,‡}, Owen N. Witte^{1,6,9,†}, Ton N. Schumacher⁵, Antoni Ribas^{1,7,8} and James R. Heath^{1,4*}

¹ NanoSystems Biology Cancer Center

² Division of Engineering and Applied Science, Bioengineering

³ Division of Physics, Mathematics and Astronomy

⁴ Division of Chemistry and Chemical Engineering, MC 127-72

California Institute of Technology, Pasadena, CA 91125

⁵ Department of Immunology, The Netherlands Cancer Institute, Plesmanlaan 121, 1066 CX
Amsterdam, The Netherlands

⁶ Department of Molecular and Medical Pharmacology, Crump Institute for Molecular Imaging

⁷ Department of Medicine, Division of Hematology-Oncology

⁸ Department of Surgery, Division of Surgical Oncology

⁹ Department of Microbiology, Immunology, and Molecular Genetics

[†] Howard Hughes Medical Institute

University of California, Los Angeles, CA 90095

[‡] Current address: Department of Chemistry, University of Illinois at Urbana-Champaign,
Urbana, IL 61801

Corresponding Author: James R. Heath

Address: 1200 E. California Blvd. MC 127-72, Pasadena, CA 91125

Phone: 626-395-6079

Fax: 626-395-2355

Email: heath@caltech.edu

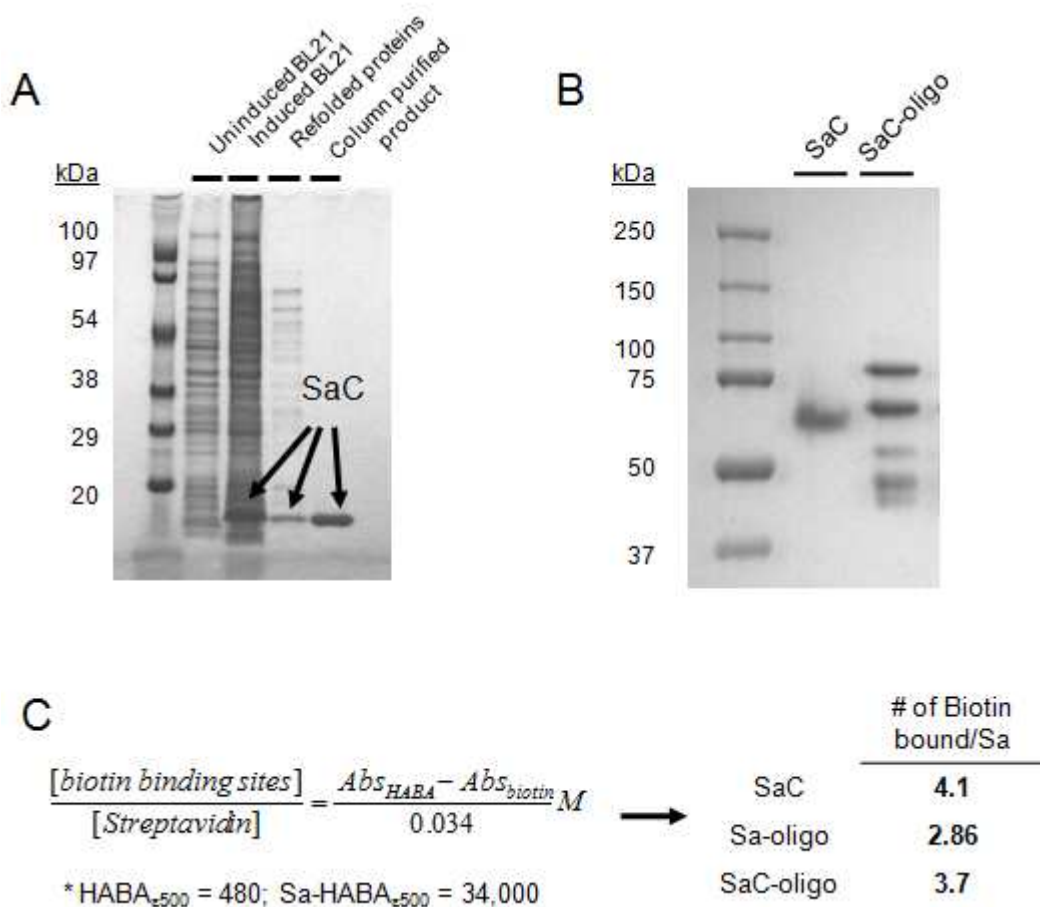


Figure S1. An engineered variant of streptavidin expressing C-terminal cysteine residues has superior biotin binding capacity compared to native streptavidin post conjugation with ssDNA. (A) The various stages of SAC expression, refolding, and purification were analyzed on a denaturing PAGE gel. The molecular weight of a SAC monomer is ~12kDa. (B) A gel mobility shift assay to verify the formation of ssDNA-SAC conjugates. Individual bands representing SAC-oligo conjugates differing by one DNA strand can be resolved. Lower order SAC-oligo conjugates (1-2 oligos per protein) run “lighter” when compared to unmodified SAC because of the difference in charge/mass density of nucleic acids. Higher order SAC-oligo conjugates corresponding to 3-4 DNA strands per SA were favored. (C) The molecule 2-(4'-Hydroxyazobenzene) benzoic acid (HABA) was used to determine the molar ratio of association of biotin to SA. Native SA-oligo conjugates bound ~2.9 moles of biotin per mole of SA, a significant decline when compared to the 4:1 ratio of unmodified SAC. SAC-oligo conjugates maintained near optimum binding capacity (3.7:1).

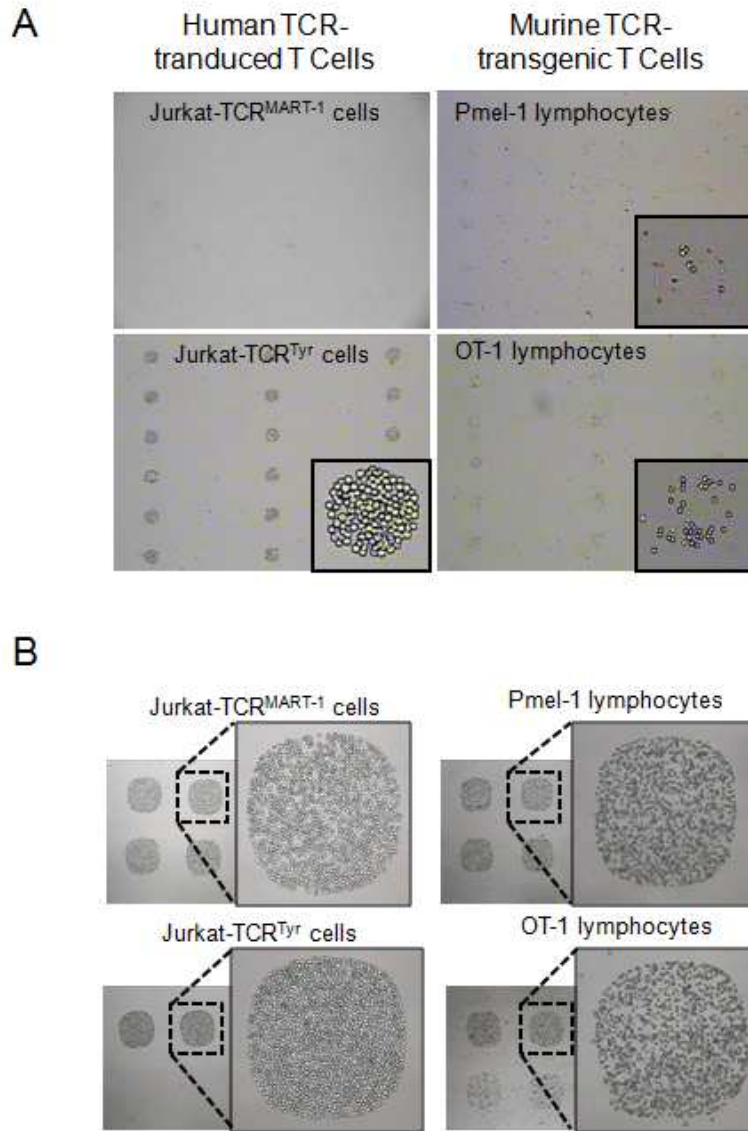


Figure S2. T cell capture efficiency is optimal when utilizing ssDNA-SAC conjugates to generate NACS p/MHC tetramers (A) ssDNA-p/MHC constructs derived from native SA were used to sort 4 different human/murine transgenic T cell populations. The T cell capture efficiencies were highly varied amongst the four T cell populations. (B) ssDNA-p/MHC tetramers derived from ssDNA-SAC conjugates were used to sort the four T cell populations. The resulting cell capture efficiencies were markedly improved over native SA-oligo conjugates, demonstrating that SAC is necessary for the production of high affinity ssDNA-p/MHC tetramers.

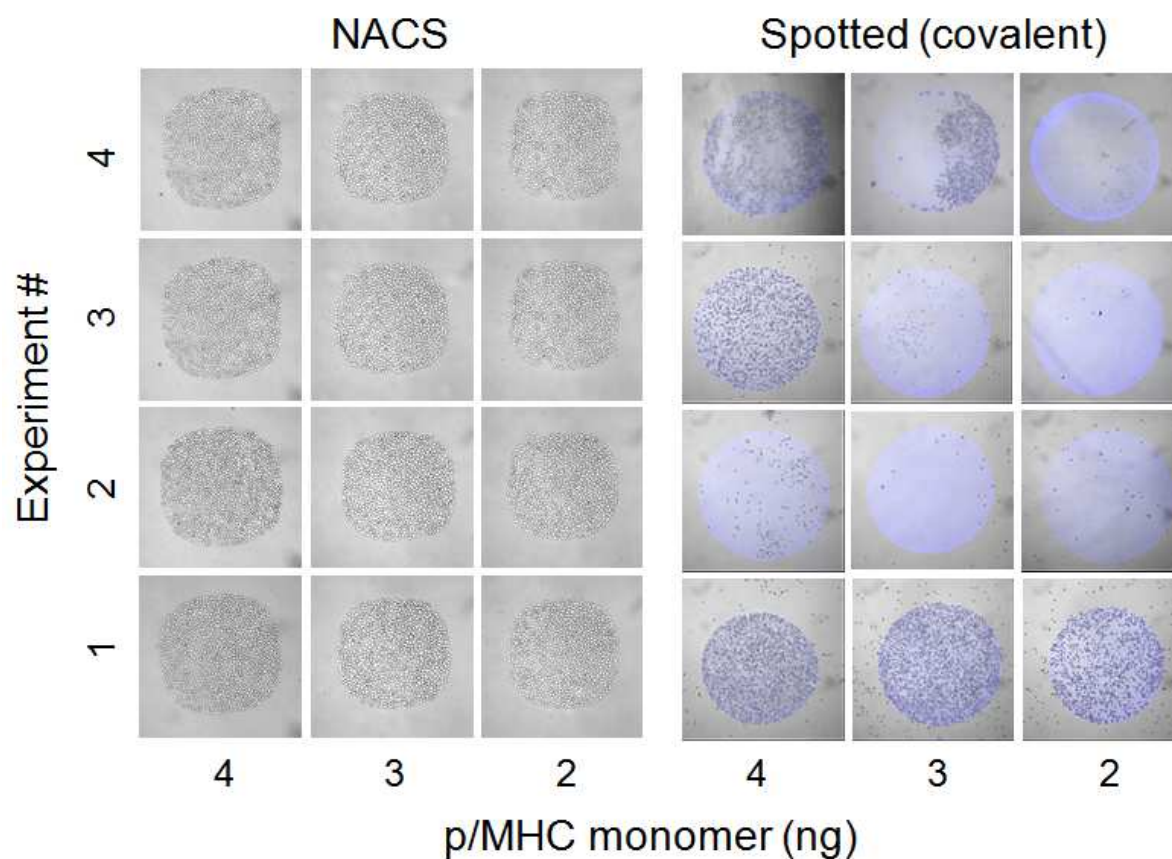


Figure S3. Comparison of the performance of p/MHC arrays produced by NACS and by spotting The consistency and robustness of T cell immobilization with NACS is evident when compared directly with spotted arrays, which suffers from significant levels of inter-spot, intra-spot, and inter-experimental heterogeneity. Each row represents a separate experiment performed on a different slide.

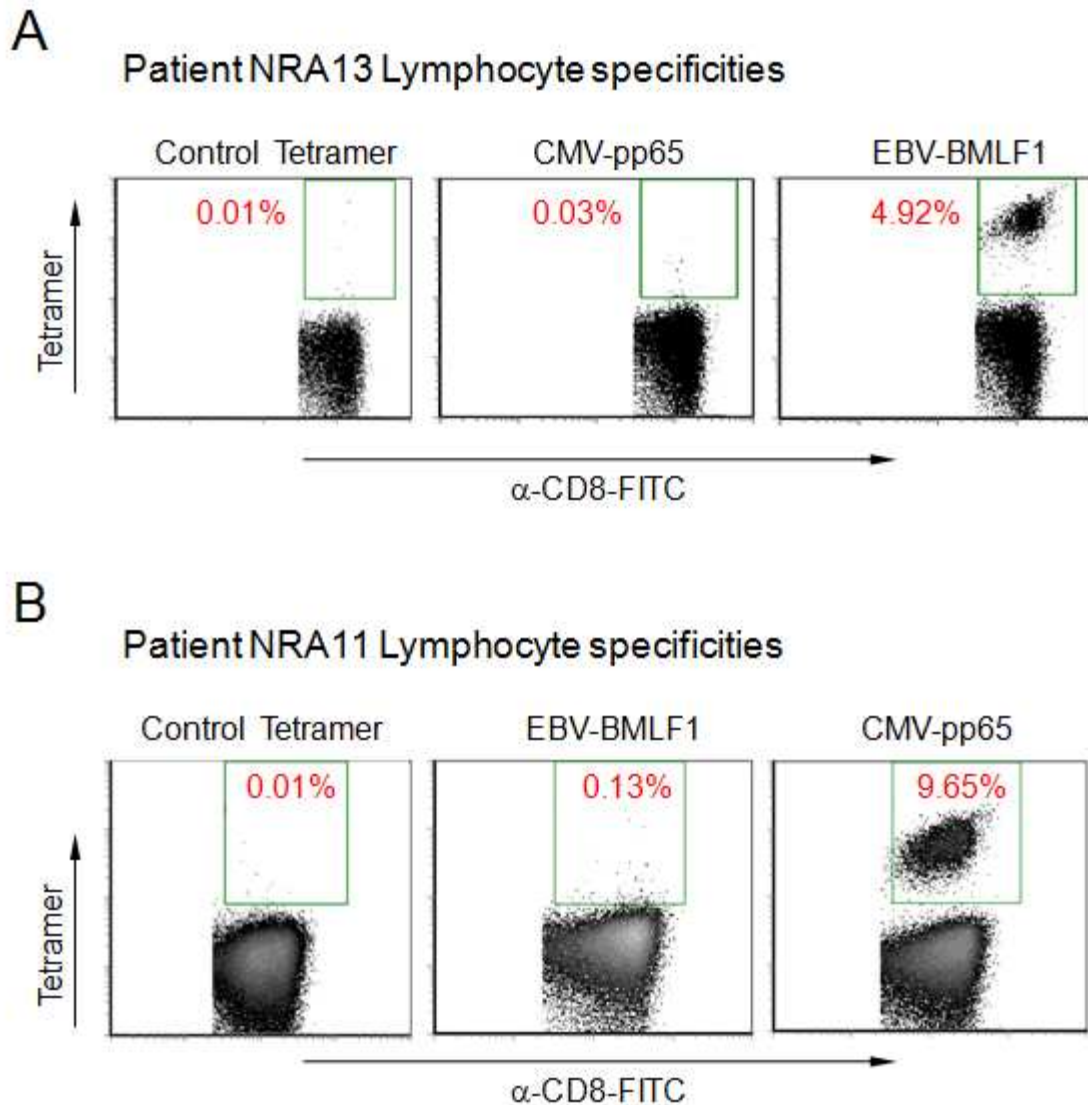


Figure S4. The T cell specificities of PBMCs from patients NRA11 and NRA13 (A)

Lymphocytes isolated from NRA13 contained significant levels of EBV specific T cells (4.9%) with minimal CMV specific T cells. (B) Lymphocytes isolated from NRA11 contained high levels of CMV specific T cells (9%) with a low population of EBV-specific cells (0.12%). Leukapheresis fractions from both patients were kept as frozen aliquots before thawing, CD8+ enrichment and NACS sorting.

Table 1. Orthogonal DNA sequences for spatial encoding of p/MHC tetramers

Name	Sequence*
A	5' - AAA AAA AAA AAA AAT CCT GGA GCT AAG TCC GTA AAA AAA AAA AAT CCT GGA GCT AAG TCC GTA AAA AAA AAA AAA A
A'	5' - NH ₂ - AAA AAA AAA ATA CGG ACT TAG CTC CAG GAT
B	5' - AAA AAA AAA AAA AGC CTC ATT GAA TCA TGC CTA AAA AAA AAA AGC CTC ATT GAA TCA TGC CTA AAA AAA AAA AAA A
B'	5' - NH ₂ - AAA AAA AAA ATA GGC ATG ATT CAA TGA GGC
C	5' - AAA AAA AAA AAA AGC ACT CGT CTA CTA TCG CTA AAA AAA AAA AGC ACT CGT CTA CTA TCG CTA AAA AAA AAA AAA A
C'	5' - NH ₂ - AAA AAA AAA ATA GCG ATA GTA GAC GAG TGC
A _{EcoRI}	5' - AAA AAA AAA AAA GAG CTA AGT CCG TAG AAT TCA AAA AAA AAA GAG CTA AGT CCG TAG AAT TCA AAA AAA AAA AAA
A _{EcoRI} '	5' - NH ₂ – AAA AAA AAA AGA ATT CTA CGG ACT TAG CTC CAG GAT
B _{BamHI}	5' - AAA AAA AAA AAA TTG AAT CAT GCC TAG GAT CCA AAA AAA AAA TTG AAT CAT GCC TAG GAT CCA AAA AAA AAA AAA
B _{BamHI} '	5' - NH ₂ – AAA AAA AAA AGG ATC CTA GGC ATG ATT CAA TGA GGC

* All sequences to be conjugated to SAC (A', B', C', A_{EcoRI}', and B_{BamHI}') were designed with a polyA linker followed by a 20mer hybridization region. The 5' amine is required for the attachment of the hetero-bifunctional maleimide derivative MHPH. Sequences printed on glass substrates (A, B, C, A_{EcoRI}, and B_{BamHI}) were designed with two hybridization regions separated by polyAs. This was designed to facilitate electrostatic adsorption to amine glass substrates.

Full References

- 39) Dudley, M.E.; Wunderlich, J.R.; Robbins, P.F.; Yang, J.C.; Hwu, P.; Schwartzentruber, D.J.; Topalian, S.L.; Sherry, R.; Restifo, N.P.; Hubicki, A.M.; Robinson, M.R.; Raffeld, M.; Duray, P.; Seipp, C.A.; Rogers-Freezer, L.; Morton, K.E.; Mavroukakis, S.A.; White, D.E.; Rosenberg, S.A. *Science* 2002, **2002**, 298, 850-854.